Elimination of Tatter Leaf-Citrange Stunt Virus from Satsuma Mandarin by Shoot-tip Grafting following Pre-heat-treatment

M. Koizumi

ABSTRACT. An excellent variety of 'Niyu' satsuma mandarin carried tatter leafcitrange stunt virus (TL-CSV). Elimination of TL-CSV from satsuma mandarin had been difficult because of its intolerance against heat treatment and failure to obtain TL-CSV-free trees by shoot-tip grafting (STG). Pre-heat-treatment of the potted Niyu satsuma at 35 C for 19-32 days or 40/30 C for 9 days plus an additional 35/30 C for 13-20 days and subsequent STG using a shoot-tip of 0.2 mm in length produced TL-CSV-free trees.

Index words. tatter leaf virus, elimination, satsuma mandarin.

Budunion crease caused by tatter leaf-citrange stunt virus (2) was recognized in several varieties or budlines grafted on Poncirus trifoliata rootstock in Japan (5, 6). In recent years certain viruses, including tatter leaf-citrange stunt virus (TL-CSV) have become widespread due to the popular practice of variety renewal by top-working. A local variety Niyu satsuma produces excellent quality fruit, but fruit set was subject to fluctuation because of TL-CSV even if the trees were approach-grafted with the resistant rootstock C. junos Sieb. ex. Tan. Elimination of TL-CSV from carrier plants was accomplished by heat treatment at 40/ (daytime/night tempera-30 C tures) for more than 60 days (3, 7, 13). However, satsuma mandarin budwood was intolerant of those temperatures (7). Shoot-tip grafting (STG) developed by Murashige et al. (8) and improved by Navarro et al. (9) was effective for elimination of viruses, viroids, and Spiroplasma citri except for TL-CSV (14, 15). This paper reports the elimination of TL-CSV from Niyu satsuma by STG following short periods of heat treatment.

MATERIALS AND METHODS

Pre-heat-treatment. A twoyear-old potted tree of Niyu satsuma grafted on rough lemon root-

This stock used. variety was carried the virus or viruses causing tatter-leaf of C. excelsa and stunting, zig-zag shoots and severe mottle of Rusk citrange, and also seedling yellows tristeza virus (CTV-SY). After removing most of leaves, the tree was held in a glasshouse at 25-30 C to force new shoot growth. When new shoots were 3-5 mm in length, the tree was transferred to a growth-chamber illuminated at 10,000 lux for 12 hours each day and the temperature controlled at 35 C (Expt. 1). In the other experiment incubation temperature was alternated from 40 C to 30 C at 12-hour intervals (Expt. 2).

Grafting. After incubation, new shoots about 5-10 mm in length were collected. Following removal of large leaves the shoots were disinfected in 0.25% sodium hypo-chlorite plus 0.1% Tween-20 and rinsed 3 times in sterile distilled water. The technique of STG was done according to Navarro et al. (10). Twelve-day-old Troyer citrange seedlings grown aseptically from seed at 28 C in the dark were decapitated 2 cm above the cotyledons. The disinfected shoot-tip was trimmed to remove leaf primodia with a razor blade sliver attached to a handle. Finally, a small shoottip consisting of the apical meristem and three to four leaf-primodia (about 0.2 to 0.4 mm in length) was excised with a razor blade sliver. The excised shoot-tip was placed on the trimmed rootstock. The grafted plant was placed aseptically on a filter-paper-platform in a test tube with roots immersed in Murashige and Skoog medium modified by Navarro et al. (9) for STG, and incubated overnight at 28 C in the dark. It was then transferred into a growthchamber illuminated at 1,000-1,500 lux for 12 hours each day at a constant 28 C. When a new sprout appeared from the grafted tissue. the intensity of the illumination was increased to 10,000 lux. Successfully grafted plants were removed from the tube and cut off at the cotyledons, when the sprout grew to 1-3 mm in length. Then they were side- or approach-grafted to a 12 year-old potted rough lemon seedling to accelerate shoot growth.

The Indexing. successfully grafted plants were indexed for presence of tatter leaf-citrange stunt virus (TL-CSV) and tristeza virus (CTV). A fully developed and mature leaf or stem was collected from each plant and the tissues were side-grafted to a rough lemon seedling, which was top-grafted with a Rusk citrange or C. excelsa scion. The indexed plant was incubated in a glasshouse at 20-24 C for 6-12 months to force shoot growth. The developing shoots were periodically observed and cut back to force new shoot growth. Indexing of CTV was done by enzyme-linked immunosorbent assay (ELISA) basically according to Clark and Adams (4) and Bar-Joseph et al. (1) using anti-CTV-SP serum.

RESULTS

The first experiment, in which a Niyu satsuma tree was incubated at 35 C and new growth subsequently removed for STG, showed that the pre-heat-treatment for 13 days was insufficient to eliminate TL-CSV from the shoot-tips of 0.2 mm in length, even if they were freed of CTV (table 1). TL-CSV in those trees caused severe symptoms such as zig-zag shoots, stunt, irregular leaves and severe mottle on Rusk citrange and tatter leaf symptoms on C. excelsa. Heattreatment for 19 days and subsequent STG produced a plant free of both TL-CSV and CTV. Four other plants were found free of CTV but not of TL-CSV. The TL-CSV within those trees produced typical symptoms of TL-CSV but less stunt on Rusk citrange. Heattreatment for 32 days and subsequent STG using shoot-tips of 0.2 mm in length produced 2 plants free of both TL-CSV and CTV.

In the second experiment, the plant was subjected to pre-heattreatment at alternate temperatures of 40 C and 30 C at 12-hourintervals. However, shoot growth of Nivu satsuma ceased completely and the sprouts gradually withered. Therefore, the incubation temperature was changed to 35/30 C after the 9-day incubation at 40/30 C. At the lower temperature regime many new sprouts developed. Excision of the shoot-tips was carefully done with lengths of 0.2 mm or less except for some tips collected from shoot the sprouts after incubation for 29 days. Pre-heat-treatment for 9 days at 40/30 C was insufficient to eliminate TL-CSV from the shoottip (table 1). Pre-heat-treatment for 9 days at 40/30 C plus an additional 6 days at 35/30 C was also insufficient. However, heat-treatment for 9 days at 40/30 C plus an additional 13 or 20 days at 35/30 C eliminated TL-CSV from shoot-tips 0.2 mm in length. But the tree grown from a 0.3-0.4 mm shoot-tip collected after heat treatment for 29 days was not free of TL-CSV. All of the successfully

Expt. no.	Pre-heat-treatment		Size of	No. of successful	No. of TL-CSV-free	No. of CTV-free
	Temp.	Period	shoot-tip	grafts	plants*	plants*
1	35 C	13 days	0.2-0.4 mm	3	0	1
	35 C	19 days	0.2 mm	5	1	5
	35 C	32 days	0.2 mm	2	2	2
2	40/30 C 40/30 C	9 days 9 days	0.2 mm	2	0	2
	+ 35/30 C 40/30 C	6 days 9 days	0.2 mm	1	0	1
	+ 35/30 C 40/30 C	13 days 9 days	0.2 mm	3	2	3
	+ 35/30 C 40/30 C	20 days 9 days	0.2 mm	3	3	3
	+ 35/30 C	20 days	0.4 mm	1	0	1

TABLE 1 RESULTS OF INDEXING FOR PRESENCE OF TATTER LEAF-CITRANGE STUNT VIRUS (TL-CSV) AND CITRUS TRISTEZA VIRUS (CTV) IN TREES OF 'NIYU' SATSUMA MANDARIN DEVELOPED BY SHOOT-TIP GRAFTING FOLLOWING PRE-HEAT-TREATMENT

*TL-CSV was indexed on Rusk citrange and C. excelsa. CTV was indexed by ELISA.

grafted plants were free of CTV. The TL-CSV carried by the trees grown from shoot-tips after heattreatment for 9-17 days produced severe symptoms when indexed on Rusk citrange. In contrast, the virus in a tree grown from the shoot-tip after heat-treatment for 29 days caused mottled leaves but no zig-zag shoots nor stunting when indexed on citrange plants.

DISCUSSION

For the elimination of TL-CSV from budwood of Meyer lemon, heat treatment for 6 weeks at 40/30 C plus an additional 2 weeks at 44/ 30 C was required, but the incubation for 16 weeks at 38 C or for 12 weeks at 40/30 C were insufficient (3). Miyakawa (7) also demonstrated that heat treatment for 90 days or more at 40/30 C was required for the elimination of TL-CSV from the budwood of certain citrus plants. TL-CSV was eliminated from Meyer lemon by preconditioning of budwood and treatment for 4-22 hours at 50 C in moist hot air (13). However, satsuma mandarin was intolerant against such heat treatment.

Roistacher et al. (14, 15) failed to eliminate TL-CSV from Meyer lemon budwood by STG and his third attempt also failed (personal communication). This experiment shows that combination of short heat-treatment and subsequent shoot-tip grafting could resolve the difficulty. Navarro et al. (11) also showed that psorosis-like pathogens could be eliminated by preconditioning plus STG. Okudai et al. (12) confirmed that pre-heattreatment for 40 days or more at 40/30 C provided CTV-free shoottips of 5-13 mm in length and they utilized an easy method of STG combined with pre-heat-treatment to obtain CTV-free plant. These indicate that the combination of pre-heat-treatment and STG is the most reliable method for elimination of virus pathogens from citrus budwood.

ACKNOWLEDGMENTS

The author is grateful to Dr. T. Miyakawa, Mr. C. N. Roistacher and Dr. S. M. Garnsey for critical review of the manuscript.

LITERATURE CITED

 BAR-JOSEPH, M., S. M. GARNSEY, D. GONSALVES, and D. E. PURCIFULL 1980. Detection of citrus tristeza virus. I. Enzyme-linked immunosorbent assay

(ELISA) and SDS-immunodiffusion methods, p. 1-8. In Proc. 8th Conf. IOCV, 1980. IOCV, Riverside.

- 2. CALAVAN, E. C., D. W. CHRISTIANSEN, and C. N. ROISTACHER
 - 1963. Symptoms associated with tatter-leaf virus infection of Troyer citrange rootstock. Plant Dis. Rep. 47: 971-75.
- 3. CALAVAN, E. C., C. N. ROISTACHER, and E. M. NAUER
 - 1972. Thermotherapy of citrus for inactivation of certain viruses. Plant Dis. Rep. 56: 976-80.
- 4. CLARK, M. F. and A. N. ADAMS

1977. Characteristics of the microplate method of enzyme-linked immunosorbent assay for the detection of plant viruses. J. Gen. Virol. 34: 475-83.
5. MIYAKAWA, T.

- 1978. A bud-union disorder of Japanese citrus on *Poncirus trifoliata* rootstock caused by tatter leaf virus. Rev. Plant Prot. Res. 11: 1-10.
- 6. MIYAKAWA, T.
 - 1980. Occurrence and varietal distribution of tatter leaf-citrange stunt virus and its effects on Japanese citrus, p. 220-24. In Proc. 8th Conf. IOCV, 1980. IOCV, Riverside.
- 7. MIYAKAWA, T.
 - 1980. Thermo-therapy for some citrus cultivars infected by tatter leaf virus. Bull. Tokushima Hort. Exp. Sta, 9: 7-11.

 MURASHIGE, T., W. P. BITTERS, T. S. RANGAN, E. M. NAUER, C. N. ROISTACHER, and P. B. HOLLIDAY 1972. A technique of shoot apex grafting and its utilization towards recovering

virus-free *Citrus* clones. HortScience 7: 118-19.

- NAVARRO, L., C. N. ROISTACHER, and T. MURASHIGE 1975. Improvement of shoot-tip grafting in vitro for virus-free citrus. J.
 - Amer. Soc. Hort. Sci. 100: 471-79.
- 10. NAVARRO, L. and J. JUAREZ
 - 1977. Elimination of citrus pathogens in propagative budwood. II. In vitro propagation. Proc. Int. Soc. Citriculture 3: 973-981.
- 11. NAVARRO, L., J. JUAREZ, J. F. BALLESTER, and J. A. PINA
 - 1980. Elimination of some citrus pathogens producing psorosis-like leaf symptoms, by shoot-tip grafting *in vitro*, p. 162-66. *In* Proc. 8th Conf. IOCV, 1980. IOCV, Riverside.
- 12. OKUDAI, N., T. TAKAHARA, and S. KUHARA
 - 1983. Obtaining virus-free citrus plants by the combined techniques of heat treatment and shoot-tip grafting. Shokubutsu boeki (Plant Protection) 37: 58-62. (in Japanese)
- ROISTACHER, C. N. and E. C. CALAVAN 1972. Heat tolerance of preconditioned citrus budwood for virus inactivation,
- p. 256-61. In Proc. 5th Conf. IOCV, 1969. Univ. Florida Press, Gainesville. 14. ROISTACHER, C. N., L. NAVARRO, and T. MURASHIGE
 - 1976. Recovery of citrus selections free of several viruses, exocortis viroid, and *Spiroplasma citri* by shoot-tip grafting *in vitro*, p. 186-93. *In* Proc. 7th Conf. IOCV, 1976. IOCV, Riverside.
- 15. ROISTACHER, C. N. and S. L. KITTO
 - 1977. Elimination of additional citrus viruses by shoot-tip grafting *in vitro*. Plant Dis. Rep. 61: 594-96.

entry, Darmited Internet, provinting

partities with Troper citrange. In this paper we describe an periments directed to describe an the characteristics of the spect of experiments inducing only electrony in Approximate and the team electrony the spect between the model interaction (recent big and the formation and Tropdescribed televa we used allowing described televa we used 2013-105 burgeon from the model and they burgeon from the model and they described televa we used 2013-105 described televa we used 2013-105 burgeon from the model is burgeon described televa we used 2013-105 burgeon from the model is burgeon described televa we used 2013-105 burgeon from the model is burgeon description the second method method method is burgeon description the second method me

REPERTNENTS AND RESIDENT

(Stress species and variation and