# **CHAPTER** 4

## **Exocortis and Related Diseases**

### **Review of Recent Research on Exocortis Disease**

#### E. O. OLSON

IN 1948, Fawcett and Klotz (8) described and gave the name exocortis to a bark-shelling disorder of *Poncirus trifoliata* (L.) Raf. rootstocks, and in 1949, Benton *et al.* (2) in Australia, demonstrated that the causal agent is a virus. Indexing by numerous investigators in many parts of the world indicated that the causal virus is widely distributed in citrus varieties. However, no obvious symptoms are produced on non-sensitive rootstocks.

The use of rootstocks sensitive to exocortis virus has increased in recent years. This is especially true since tristeza-tolerant rootstocks such as trifoliate orange, Rangpur lime (*Citrus reticulata* var. *austera* hyb.), and Troyer citrange (*P. trifoliata* x *C. sinensis*) have been substituted for sour orange (*C. aurantium* L.) rootstock in areas where tristeza occurs. When budwood infected with exocortis virus is propagated on these rootstocks, bark shelling, retarded growth, and reduced yields result. In many areas considerable losses have resulted because propagators ignored the general but fundamental rule of propagating only budwood from vigorous, productive, long-lived trees growing on the same rootstock as that used for propagation.

The effects of exocortis virus on tolerant rootstocks are more subtle. For example, grafts of lemon [C. limon (L.) Burm. f.] infected with

#### EXOCORTIS and RELATED DISEASES

exocortis, but free of other citrus viruses, caused measurable stunting of trees on the "tolerant" rootstocks: sweet orange, grapefruit (C. paradisi Macf.), and sour orange (5). Exocortis-infected Washington navel orange trees on Cleopatra mandarin (C. reticulata Blanco) and sweet orange rootstocks showed no obvious bark symptoms, but grew more slowly than virus-free trees, and infected trees on sweet orange rootstock produced less fruit than did virus-free trees (20).

Exocortis virus also affects sweet lime [C. aurantifolia (Christm.) Swing.], sweet lemon [C. limon (L.) Burm. f.], Cuban shaddock, possibly a lemon-citron hybrid, and Tahiti lime (15, 17, 23).

#### Indexing for Exocortis

Since exocortis virus causes no obvious symptoms on many citrus varieties, its presence in symptomless carriers may be determined only by indexing, i.e., the procedure of grafting tissue of the suspected host onto sensitive indicator plants. Likewise, failure to transmit exocortis virus by grafting tissue of a tree to the most sensitive indicator is the best proof that the tree under test is free of exocortis virus.

Benton et al. (2) in 1949, in Australia, were indexing sweet orange selections when they showed that exocortis is graft-transmissible and that symptoms appear 4 to 8 years after infected sweet orange tissue was budded onto *P. trifoliata*. Moreira (12) in Brazil, and Olson and Shull (14) in the United States, showed that exocortis virus caused bark shelling of Rangpur lime within 44 months after budding. Thus, Rangpur lime provided a more rapid test for exocortis virus than did *P. trifoliata*. Childs et al. (7), in the United States, developed a test for infection based upon histochemical examination of phloem ray cells of *P. trifoliata*. It too was replaced by newer tests.

Moreira (13) showed that yellow blotching of Rangpur lime and P. trifoliata bark was a symptom of exocortis infection and that it occurred 4 to 6 months after infection in vigorous plants. In some areas blotching from unidentified causes reduces the reliability of this test.

In the United States, Calavan and Weathers (3) included citron (C. *medica* L.) among a list of species with symptoms resembling those produced by exocortis. In Brazil, Salibe (16) showed that some varieties of citron reacted severely to exocortis infection within 200 days. In 1964, a team of Californians (4, 10) developed an indexing procedure based on selected seedlings of Etrog citron. Leaf symptoms of exocortis infection developed in the selections five to ten weeks after inoculation. This test is currently the best one for indexing exocortis virus in possible bud-

#### PROCEEDINGS of the IOCV

wood sources. Thus, in the period 1949 to 1964, the time required to index for exocortis virus was reduced from eight years to ten weeks. This development resulted from cooperation and exchange of data among research workers in Australia, Brazil, and the United States.

Different strains of exocortis (6, 18, 23) exist, and some strains stunt trees in the absence of bark-scaling symptoms (9). Strains of exocortis virus that cause no recognizable symptoms on Rangpur lime and *P. tri-foliata* are detected by the citron test (4).

Plant nutrition also affects the effectiveness of indexing methods. Weathers *et al.* (24) obtained better symptom development and shorter incubation periods on *P. trifoliata* plants by using high levels of nitrogen and phosphate fertilizer.

### Virus Spread

Reports by many observers suggest that exocortis virus is spread mainly through propagation of infected plants. However, Weathers (21, 22) transmitted the virus experimentally from citrus through dodder (*Cuscuta subinclusa*) to citrus and *Petunia hybrida*. He also transmitted exocortis from petunia to petunia by grafting and by mechanical transfer in sap. Natural spread from citrus to citrus in the greenhouse has been reported (4). No evidence of seed transmission of exocortis has appeared in seedlings from infected trees in Australia (9), South Africa (1), and California. However mild strains of exocortis occurred in noninoculated nucellar seedlings of exocortis-infected Baianinha navel orange in Brazil (19). Since this review was presented in 1966, Garnsey and Jones (11) have shown that exocortis virus was transmitted mechanically on grafting tools to 26 of 30 plants in three tests.

#### Current Problems

The principal unanswered questions affecting the problem of exocortis virus infection are as follows: In what way do weak strains of exocortis affect citrus varieties on different commercial rootstocks? Does seed transmission of exocortis virus occur? Does a virus other than exocortis virus cause stunting of trees on *P. trifoliata* rootstock? How do variations in environment affect expression of symptoms?

#### Literature Cited

- BASSON, W. J., and SCHWARZ, R. E. 1964. Indexing for exocortis and xyloporosis in mother trees of a collection of citrus cultivars. S. African J. Agr. Sci. 7: 627-632.
- 2. BENTON, R. J., BOWMAN, F. T., FRASER, L., and KEBBY, R. G. 1949. Stunting

94

and scaly butt of citrus associated with *Poncirus trifoliata* rootstock. Agr. Gaz. N. S. Wales 61: 521-526, 577-582, 641-645, 654.

- CALAVAN, E. C., and WEATHERS, L. G. 1959. The distribution of exocortis virus in California citrus, p. 151-153. In J. M. Wallace [ed.], Citrus Virus Diseases. Univ. Calif. Div. Agr. Sci., Berkeley.
- CALAVAN, E. C., FROLICH, E. F., CARPENTER, J. B., ROISTACHER, C. N., and CHRISTIANSEN, D. W. 1964. Rapid indexing for exocortis of citrus. Phytopathology 54: 1359-1362.
- 5. CALAVAN, E. C., and WEATHERS, L. G. 1959. Transmission of a growth-retarding factor in Eureka lemon trees, p. 167-177. In J. M. Wallace [ed.], Citrus Virus Diseases. Univ. Calif. Div. Agr. Sci., Berkeley.
- CALAVAN, E. C., and WEATHERS, L. G. 1961. Evidence for strain differences and stunting with exocortis virus, p. 26-33. In W. C. Price [ed.], Proc. 2nd Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- CHILDS, J. F. L., NORMAN, G. G., and EICHHORN, J. L. 1958. A color test for exocortis infection in *Poncirus trifoliata*. Phytopathology 48: 426-432.
- 8. FAWCETT, H. S., and KLOTZ, L. J. 1948. Exocortis of trifoliate orange. Citrus Leaves 28(4): 8.
- FRASER, L. R., LEVITT, E. C., and Cox, J. 1961. Relationship between exocortis and stunting of citrus varieties on *Poncirus trifoliata* rootstock, p. 34-39. *In* W. C. Price [ed.], Proc. 2nd Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- FROLICH, E. F., CALAVAN, E. C., CARPENTER, J. B., CHRISTIANSEN, D. W., and ROISTACHER, C. N. 1965. Differences in response of citron selections to exocortis virus infection, p. 113-118. In W. C. Price [ed.], Proc. 3d Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- GARNSEY, S. M., and JONES, J. W. 1967. Mechanical transmission of exocortis virus with contaminated budding tools. Plant Disease Reptr. 51: 410-413.
- MOREIRA, SYLVIO. 1955. A molestia "exocortis" e o cavalo de limoeiro cravo. Revista de Agricultura (Piracicaba) 30: 99-112.
- MOREIRA, S. 1961. A quick field test for exocortis, p. 40-42. In W. C. Price [ed.], Proc. 2nd Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- OLSON, E. O., and SHULL, A. V. 1956. Exocortis and xyloporosis-bud-transmission virus diseases of Rangpur and other mandarin-lime rootstocks. Plant Disease Reptr. 40: 939-946.
- OLSON, E. O., SHULL, A., and BUFFINCTON, G. 1961. Evaluation of indicators for xyloporosis and exocortis in Texas, p. 159-165. In W. C. Price [ed.], Proc. 2nd Corf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- SALIBE, A. A. 1961. Contribuição ao estudo da doenca exocorte dos citros. 71 p. Mimeographed. Doctoral thesis. Univ. de São Paulo, Brazil.
- SALIBE, A., and MOREIRA, S. 1965. Tahiti lime bark disease is caused by exocortis virus, p. 143-147. In W. C. Price [ed.], Proc. 3d Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- SALIBE, A., and MOREIRA, S. 1965. Strains of exocortis virus, p. 108-112. In W. C. Price [ed.], Proc. 3d. Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- SALIBE, A., and MOREIRA, S. 1965. Seed transmission of exocortis virus, p. 139-142. In W. C. Price [ed.], Proc. 3d Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.
- SINCLAIR, J. B., and BROWN, R. T. 1960. Effect of exocortis disease on four citrus rootstocks. Plant Disease Reptr. 44: 180-183.

- WEATHERS, L. G. 1965. Transmission of exocortis virus of citrus by Cuscuta subinclusa. Plant Disease Reptr. 49: 189-190.
- 22. WEATHERS, L. G. 1965. Petunia, an herbaceous host of exocortis virus of citrus. (Abstr.) Phytopathology 55: 1081.
- 23. WEATHERS, L. G., and CALAVAN, E. C. 1961. Additional indicator plants for exocortis and evidence for strain differences in the virus. Phytopathology 51: 262-264.
- WEATHERS, L. G., HARJUNG, M. K., and PLATT, R. G. 1965. Some effects of host nutrition on symptoms of exocortis, p. 102-107. *In* W. C. Price [ed.], Proc. 3d Conf. Intern. Organization Citrus Virol. Univ. Florida Press, Gainesville.